

luminescence - emission of light by substance when a phosphorescent → delayed fluorescence. is phot.

Fluorometry

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Fluorescence

↳ when molecule absorb light at one λ and emit light at a longer λ .

↓
atom that fluoresces → fluorophore

measurement of emitted ^{fluorescent} light = fluorometry

- light entering sample is not measured, so stray light effect is reduced. → accurate and sensitive

Principle :-

Molecule contain series of closely spaced energy level

↓
when light energy is passed on molecule

↓
Absorptⁿ of light energy by molecule

↓
Transition of electron from ground state to one of excited state

↓
Once excited, e^- returns to original ground state by several ways

radiation-less
transⁿ equilibrium

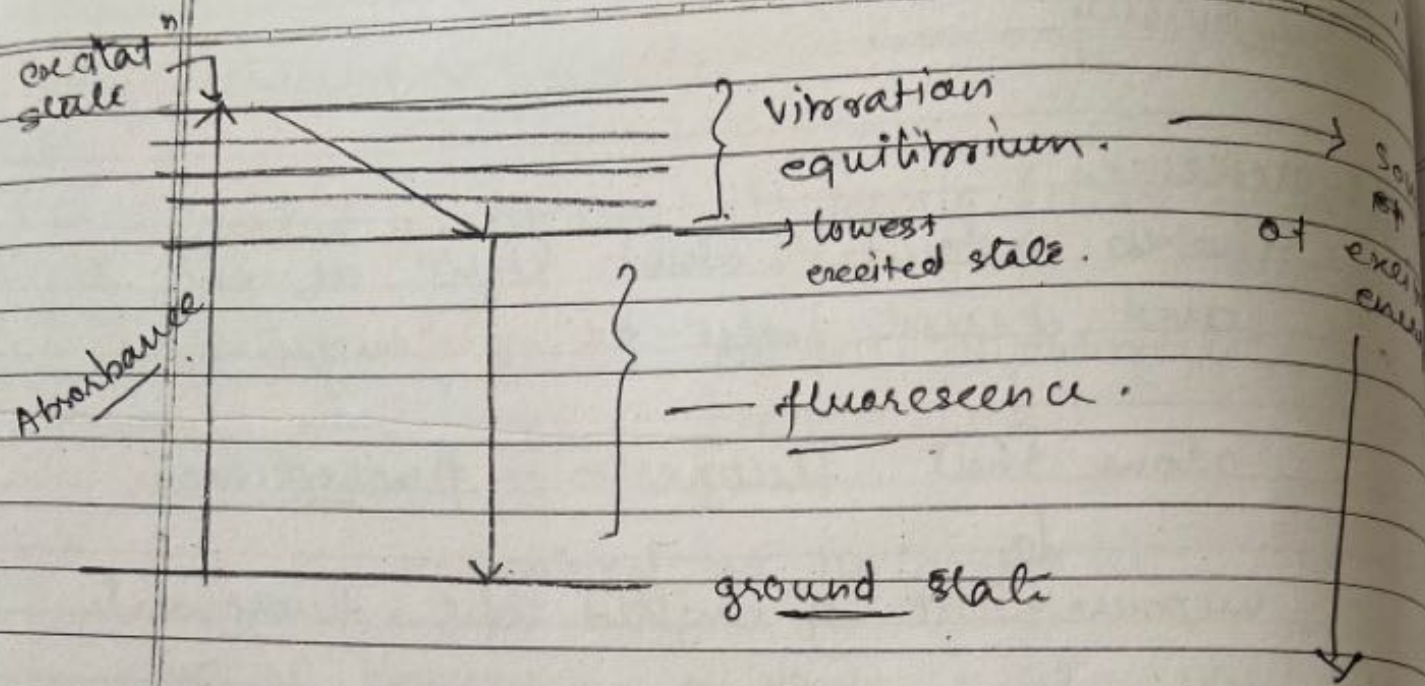
↓
emission of visible light

quenching

↓
trapping of energy.

↓
fluorescence

↓
as heat



Thus, emitted fluorescent light is of less energy (higher λ) than excitation energy.

↓
 Difference b/w λ of ^{max} excitation ~~exc~~ light and emitted light

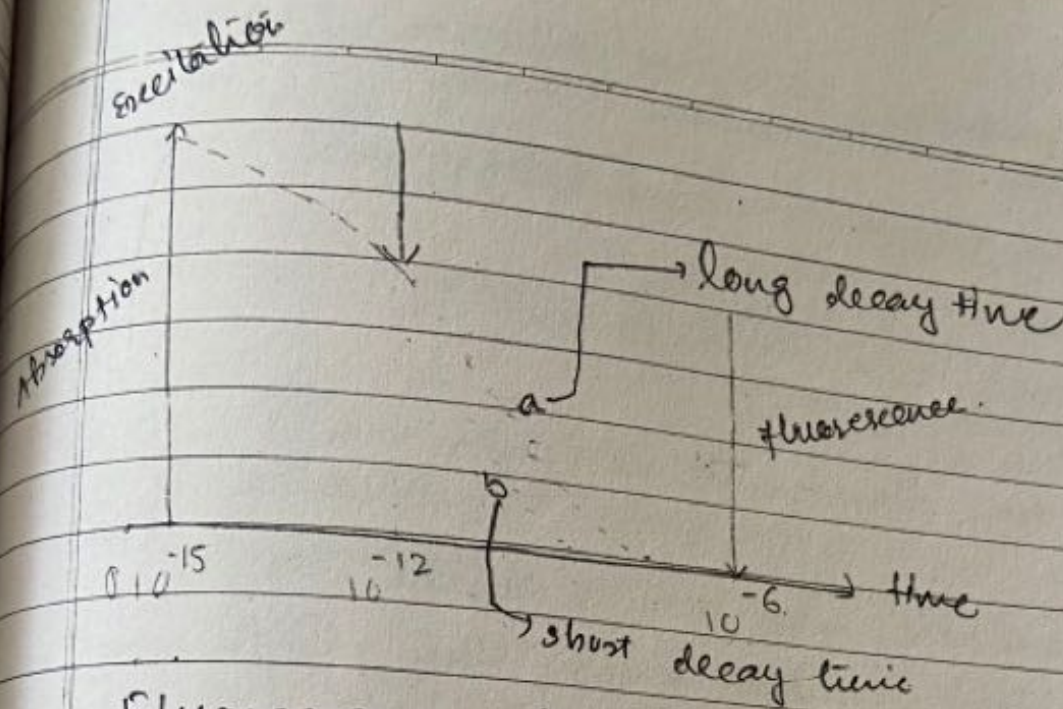
↓
Stokes shift, (constant)
 (measure of loss of energy during lifetime of excited state (vibration equilibrium) before returning to ground state..)

⇒ Time relationship of fluorescence emission.

There is considerable delay time between the

- ① Absorption of light energy
- ② Vibration equilibrium.
- ③ emission of fluorescence. See behind.

↓
 shown in figure.



Fluorescence decay process.

→ Relationship of concentration and fluorescence intensity

• Fluorescence intensity directly \propto to concentration of fluorophore.

- Magnitude of intensity determined by

- ① Conc.
- ② length path
- ③ Intensity of light source.

• Fluorescence measurement is 100-2000 times more sensitive than abs. measurement

- dit
- ① more intense light
 - ② Digital filtering tech.
 - ③ Sensitive emission photometer.

• Time relationship of fluorescence emission

• Time req. for molecule to absorb energy and to be excited = 10^{-15} sec

• length of vibratⁿ equilibrium to lowest excited state = 10^{-12} sec.

• time req. for fluorescence emission = 10^{-7}

→ There is considerable ^{time} delay b/w ① Absorbⁿ of energy ② return to lowest energy state ③ flou. emissi-

→ Fluorescence decay time = time required for the emitted light to reach $1/e$ where $e = 2.718$ (Napierian base) at its initial intensity is called fluorescence decay time / average life time of excited state molecule.

↓
this time delay is used in fluorescence instrument → time resolved fluorescence

↓
pulse of light (Xenon lamp / laser) is used

↓
fluorescence is measured ⁱⁿ time following pulse of light.

Advantage → ① ↓ effect of stray light and
② no light scattering of incident light
↓ background interference.

② Monochromator :- ^{excitation} emission.

• Interference filter

- all-dielectric multicavity filter
- combine \bar{c} sharp cut off filter \rightarrow single filter package

- remove undesired transmission.
- provide narrow bandwidth, higher peak transmission

• Coloured glass.

- used for both excitation & emission filter.
- max chances of stray light transmission \downarrow unwanted fluorescence.

- Coating \rightarrow provide selectivity of excitation and emission λ required when work with new fluorophore.

③ Cuvets

- square / rectangular.
- material allows excitation and emission light to pass

- glass / plastic - for visible light.
- quartz - UV light

* Relation of concentration and fluorescence intensity derived from Beer's law. Lambert's law.

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fluorescence intensity of concentration & intensity of incident light

↓
↑ concentration } ↑ fluorescence.
↑ lamp intensity }

→ Inner filter effects

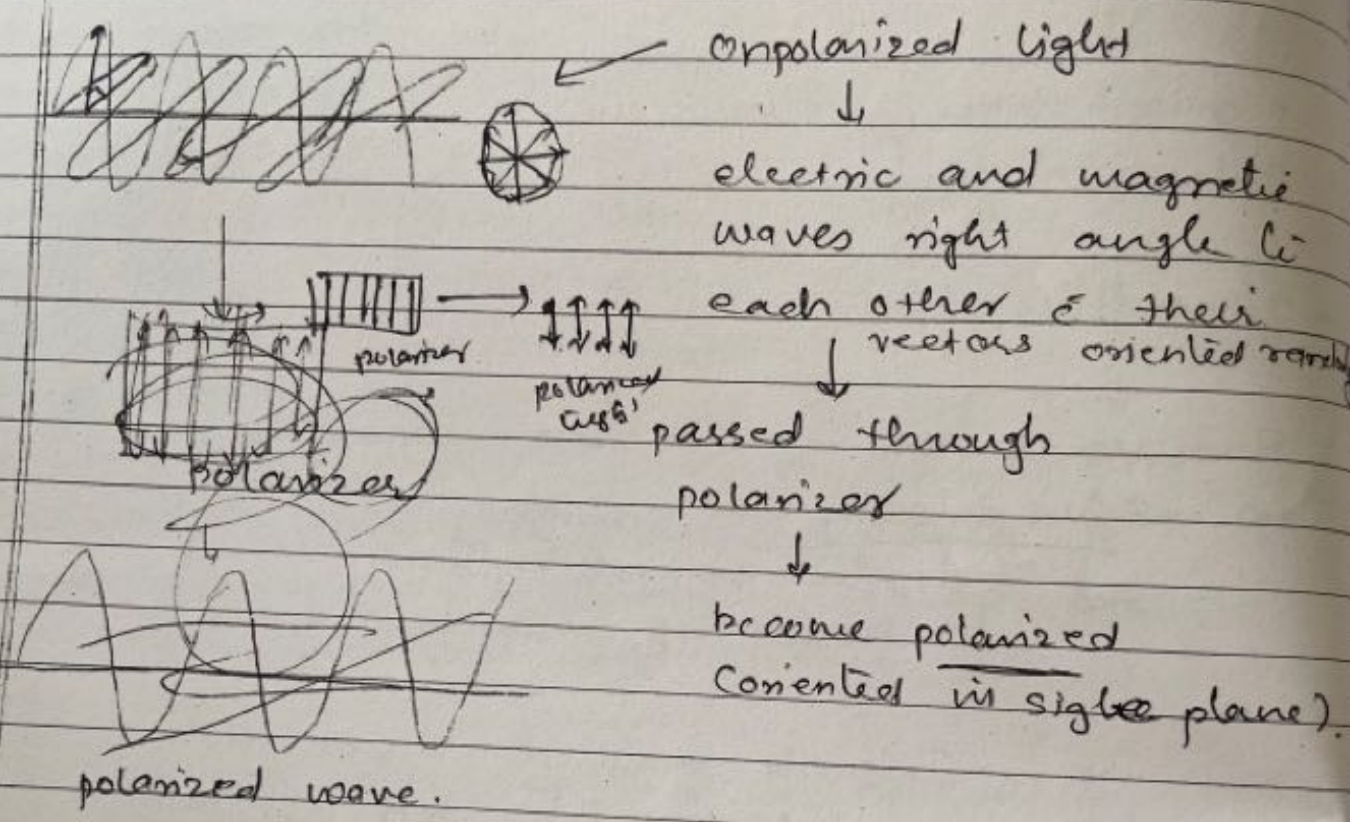
→ Fluorescence measurement are expressed in relative intensity unit which are ^{arbitrary} ~~arbitrary~~ unit decided by manufacturer.

↓
Intensity measured is not the absolute quantity, but a small part of total fluorescence emission

↓
its magnitude is defined by instrument slit width, excitation intensity & detector sensitivity (instrument-related variables)

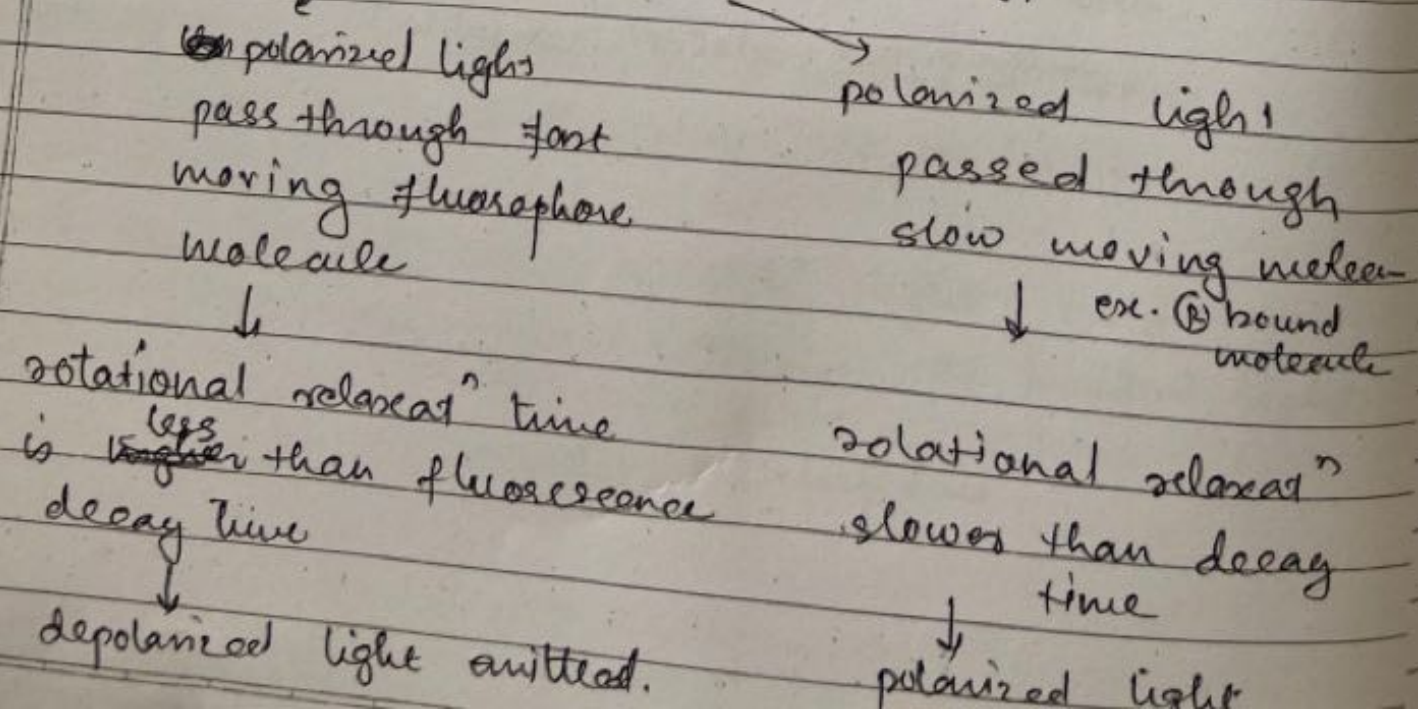
* Fluorescence polarization :-

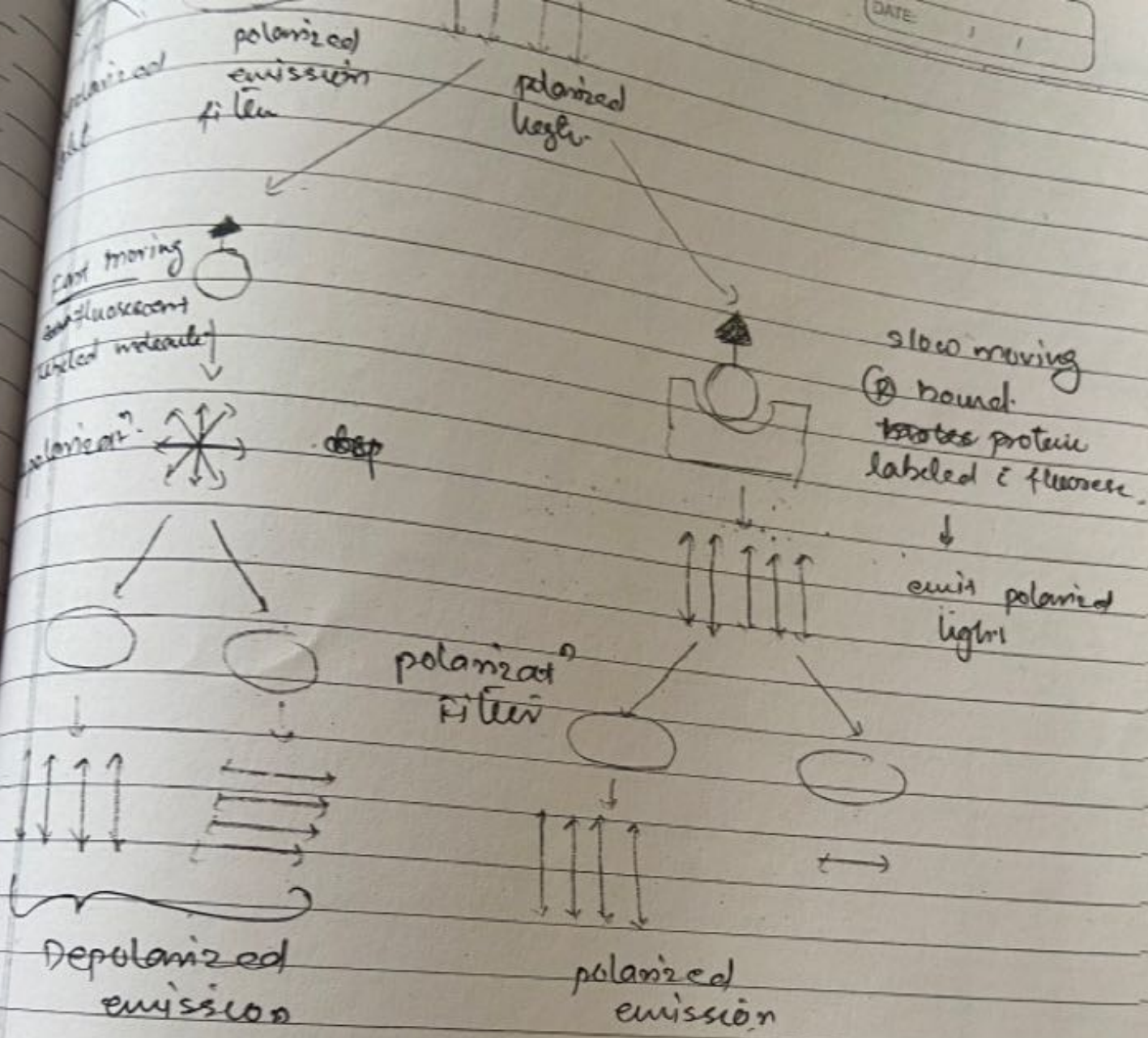
→ light is composed of electric and magnetic waves at right angle to each other.



unpolarized light
 For sub-label
 depolar

→ Fluorophore absorbs light most efficiently in plane of electronic energy.

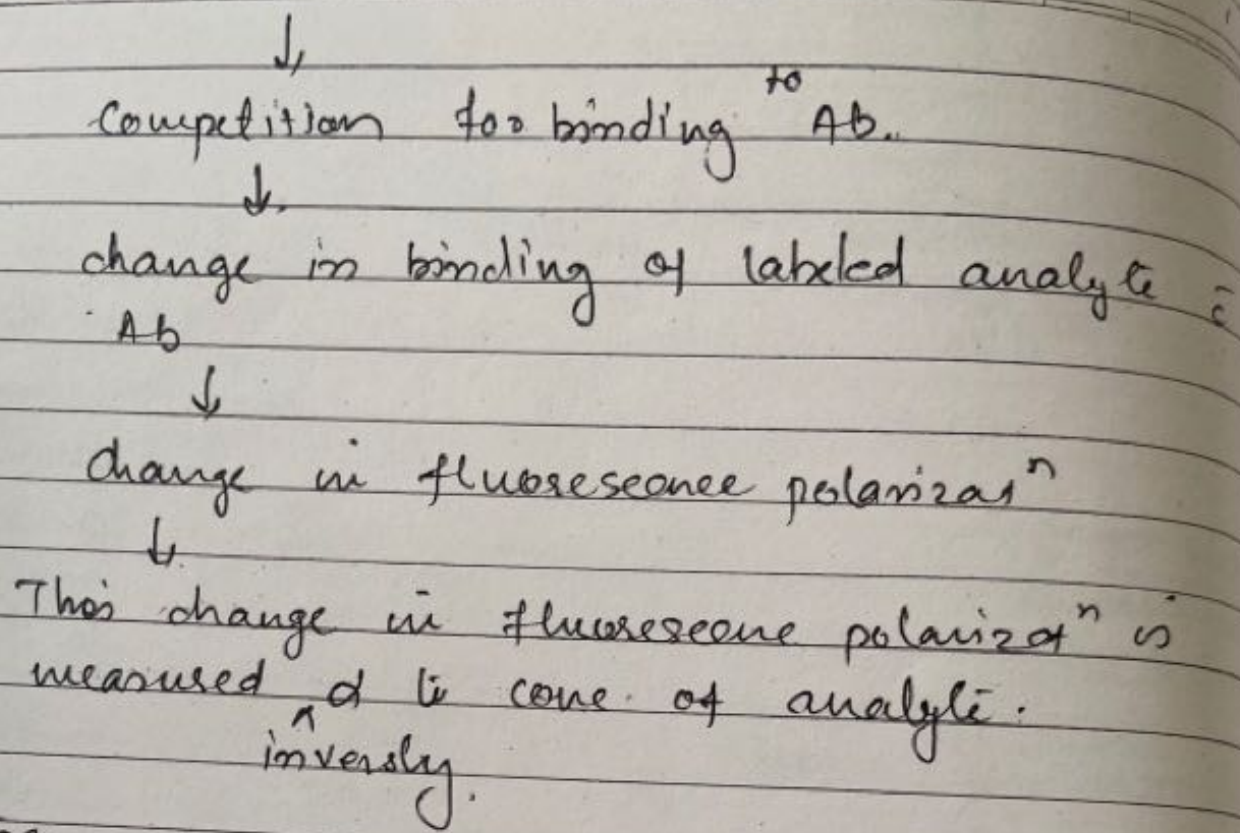




Use of fluorescence polarization :-

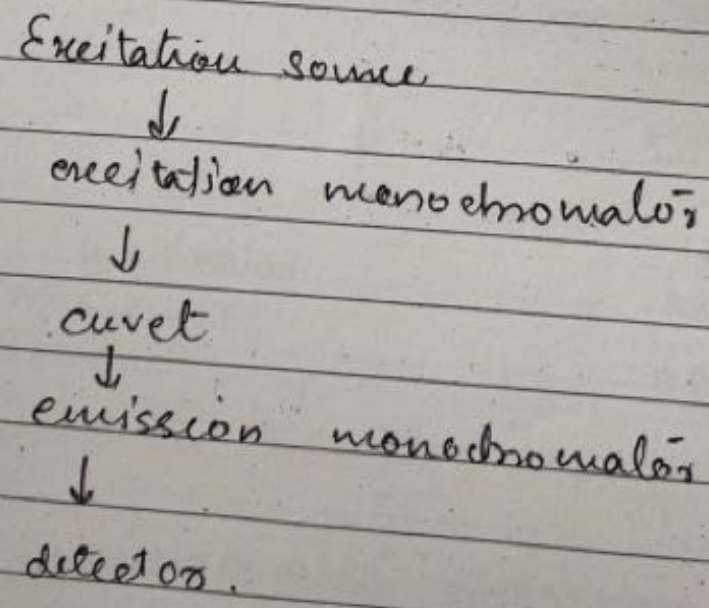
- ↓
- quantitative analysis by change in fluorescence polarization of immunological reaction
- ↓
- labeled Ab Analyte bind to Ab
- ↓
- emission of polarized light
- ↓
- Addition of nonlabeled molecules e.g. drugs in serum sample.
- ↓

2. ATP bind and hydrolyse to change pro



• Advantage - Homogenous immunoassay → Automated
↓
↓

↓ Instrumentation



① Excitation sources:-

- Absorption spectra of most of fluorophore = 300-700 nm

- Fluorescence emission of

- ① initial excitatⁿ energy
- ② Conc. of analyte

↓
Intense lamp capable of emitting radiant energy of over large spectrum desirable

ex → ① X-ray lamp

② Mercury arc lamp

③ Laser

(i) X-ray lamp - not used.

- high intensity radiant energy → 20 to 500 nm
- stable lamp flashes.
- high UV and visible spectrum output.

Disadvantage → are wandering and flickering
by fluctuatⁿ in current

↓
prevented by current-stabilized power supply

(ii) Laser:-

used in fluorescence applicatⁿ where high intensity, well focused and monochromatic light is req.

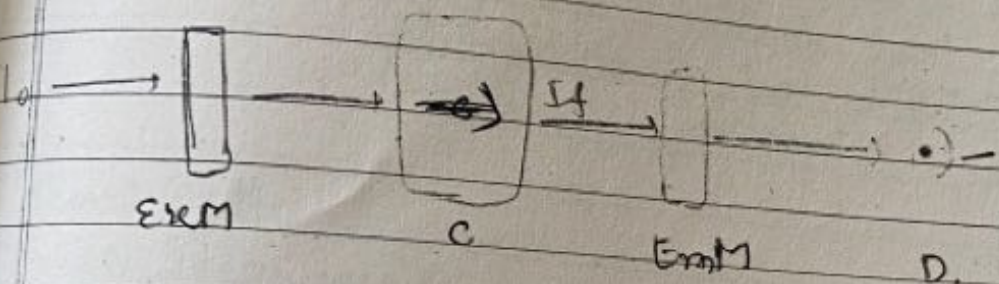
applicatⁿ - time resolved
flow cytometry

Geometric arrangements of fluorimeters :-

Fluorescence light is emitted in all directions from molecule

↓
several geometry are used to measure fluorescence.

① End-on geometry



EMM = Excitation monochromator

C = Cuvet

EmM = Emission

D = detector

I_0 = initial excitation energy

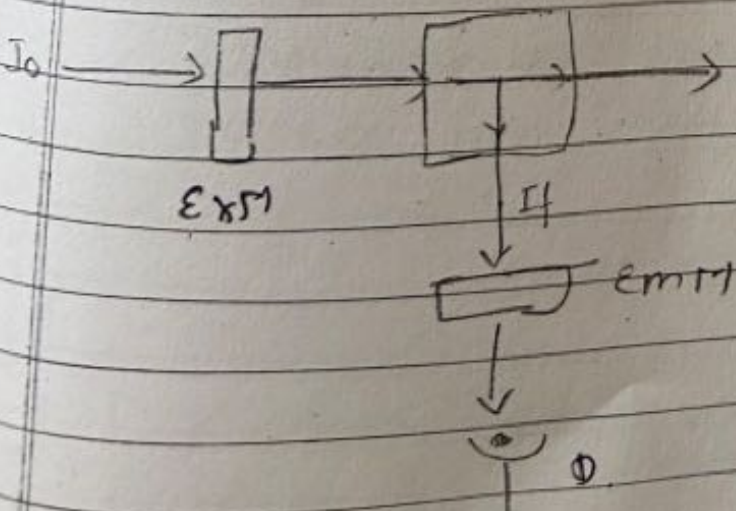
I_f = fluorescence energy

- Not used widely
- limited sensitivity due to

- ① quality of excitation and emission filter pair
- ② overlapping of ex and em spectral band
- ③ Inner filter effect.

• Advantage - can be combined in spectrophotometer

② Right angle geometry



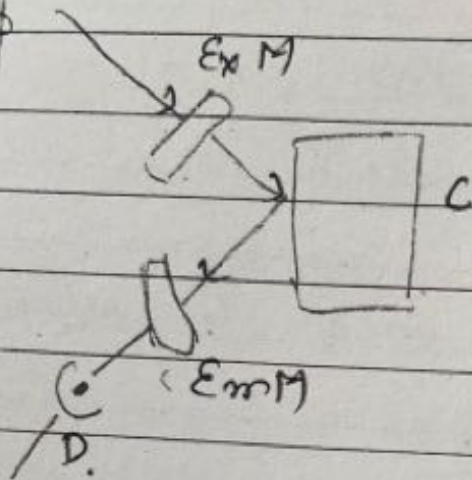
• Most commonly used

• ↓ background signals
↓ scattering.

↑ Analytical sensitivity

Disadv = high inner filter effect

③ front-surface geometry

I₀

- minimise inner effect

↓

↑ linearity over range of cone.

- Similar sensitivity to right angle, but more susceptible to background light scattering.

Limitation of fluorescence measurement!

Factors

- ① Concentration effects → inner filter effect
→ concentration quenching
- ② Background effects → ditto
- ③ Solvent effects → scattering
→ Interfering non specific fluorescence, or quenching from solvent
- ④ Sample effect → light scattering
→ Interfering fluorescence
→ Sample adsorption
- ⑤ temp. effects
- ⑥ photo decomposition of sample

① Inner filter effects:-

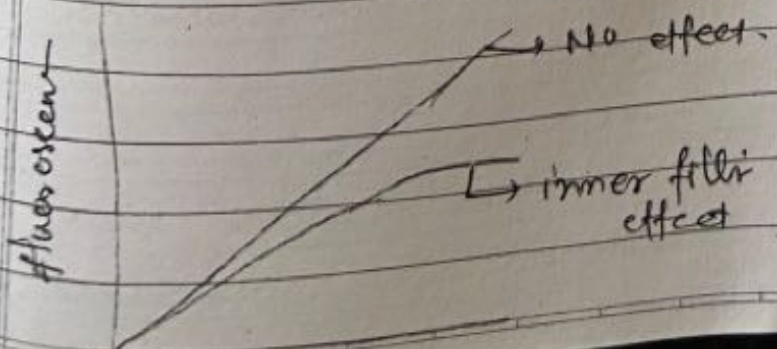
fluorescence intensity in the sample

fluorophore itself and other non-fluorescent molecules absorb exciting light or emission light

↓
loss of excitation intensity across the cuvet path length

↓
relationship b/w concentration and fluorescence becomes non linear (if abs. > 2%)

↓
problem is more common in right angle fluorescence emission slit is set at 90° to center of sample cell.



↑ absorption

→ problem solution

①

① use of front-surface fluorescence instrument

↓

emission slit is located near the front of sample cell.

② Dilution of Solution

③ Concentration quenching

(i) Detectⁿ of macro-
molecules eg Ab

↓

heavily labeled
fluorophore.

○

or

(ii) in flow cytometry /
laser induced fluorescence

↓

where

↑ red density of fluorophore
label.

↘ ✓
when excited, fluorescence labels
are in such a close proximity

↓

radiation-less energy transfer

↓

quenching

↓

Resulting fluorescence is much lower
than expected.

slight blue color of Rayleigh scattering of short rays \rightarrow blue by molecules having size less than λ

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③ Light scatterings -

↓
Rayleigh scattering

↓
Raman scattering

• scattering \bar{c} change in λ

• \bar{c} lengthening of wavelength

• more common in

• Independent of excitation and it's a property of solvent

fluorophores \bar{c} excitation emission spectra overlap (small Stokes shift)

↓
minimized by use of excitation and emission interference filter or use of polarizer.

↓
minimized by setting excitation and emission λ far enough to prevent Raman scattering. $\&$ narrowing slit width.

(1) Cuvet material and solvent effects:-

certain Cuvet materials - quartz glass / plastic

↓
contain UV absorbers

↓
give fluorescence.

certain solvent \rightarrow eg. ethanol

↓
give fluorescence.

• Cuvet and solvent \bar{c} minimal fluorescence emission are commercially available.
 \hookrightarrow ↓ background fluorescence